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# Prevalence Of Methicillin- Resistant Staphylococcus Aureus And Coagulase-Negative Staphylococci Among Male Students In A Private Tertiary Institution And Their Enterotoxin -producing Potential

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ARTICLE HISTORY		ABSTRACT	
Received:	13-Jun-2011	In order to determine the prevalence and antimicrobial resistance among methicillin-resistant, methicillin-sensitive and coagulase-	
Accepted:	11-Aug-2011	negative staphylococci in a tertiary institution, nasal swabs were collected from 100 male students. A total of 98 staphylococci were	
Available online	:: 10-Feb-2012	isolated, out of which 51 were coagulase positive. Methicillir resistance among the <i>Staphylococcus aureus</i> isolates was 25.5% The carriage rate of methicillin-resistant coagulase-negative staphylococci among the subjects was 11%. <i>Bacillus</i> spp. was isolated in pure cultures from 5 subjects and of these isolates one was	
Methicillin resistance, staphylococci, enterotoxins, tertiary institution		resistant to methicillin. Antimicrobial susceptibility of the methicillin- resistant <i>Staphylococcus aureus</i> (MRSA) are methicillin-resistant coagulase negative staphylococci (MRCONS isolates to various antibiotics tested showed that, in most cases, the MRSA were less susceptible to the antibiotics than MRCONS. Not of the MRSA was resistant to vancomycin. There were no identified risk factors in this study. Twenty-six strains of <i>Staphylococcu aureus</i> isolates produced staphylococcal enterotoxin	
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E-mail: mosola	toye@yahoo.com	from the 13 MRSA isolates). <i>Staphylococcus saprophyticus</i> produced SEA(11 strains), SEB (2 strains) and SEC (2 strains).	

# **INTRODUCTION**

*Ctaphylococcus aureus* infections have continued to rise in Dhealth care facilities in recent times, with increased emergence of strains resistant to methicillin called methicillinresistant S. aureus (MRSA). Studies have also shown that these strains have become important in community-acquired infections [1-4]. A number of studies have investigated health care workers and individuals associated with them for nasal carriage of S. aureus. Weng-Tsung et al. [5] and Ciftci et al. [6], in their studies involving children in Turkey and Taiwan reported 13.2% and 0.3% community-acquired MRSA carriage in health workers' children respectively. In a similar study involving adult health care workers', Kalsoom et al. (7) found the prevalence of nasal carriage of S. aureus, coagulase-negative staphylococci (CONS) and MRSA to be 48%, 46% and 14% respectively. A 2007 report estimated the number of MRSA infections treated in hospitals to have doubled in the USA (8). Many clinical infections arise from spread from healthy carriers but most surveillance of S. aureus and MRSA have focused on individual with invasive infection rather than on an entire population (4,9). S. aureus nasal carriage, present in about 20% of the general population, has been

identified as a key factor for the subsequent development of community-acquired and nosocomial staphylococcal infections (10, 11). Methicillin-resistant S. aureus carriage has been significantly correlated with the presence of skin lesions, antibiotic usage and prior hospitalization (12). There is paucity of data on the prevalence and antibiotic susceptibility patterns of MRSA in community settings such as a University far removed from a hospital setting. Most data on the prevalence and susceptibility patterns of S. aureus in Nigeria have been from clinical isolates. Nasal carriage of methicillin-resistant S. aureus has varied prevalence from region to region and from one country to another (13, 14). Proper knowledge of the prevalence and local antimicrobial resistance patterns of S. aureus is essential for understanding the epidemiology of S. aureus infections and prescription of adequate therapy and formulation of infection control policies. There are few reports in Nigeria on the enterotoxin production ability of isolates of S. aureus from nasal passage of healthy carriers. This study aims at documenting the prevalence, antibiotic susceptibility patterns and enterotoxigenicity potentials of staphylococci isolated from male students in a private University in southwestern Nigeria.

# **MATERIALS AND METHODS**

#### Sample population, Specimen Collection & Processing

The study population involves male students of Redeemer's University, Mowe, Ogun State, Nigeria. Informed consent was obtained from the students before samples collection. The sampled population was about 12% of the total male students in the University at the time when this study was carried out. All participants were students who had not been exposed to the health care system or attended hospitals or used any antibiotic during the previous 6 months. Written questionnaires containing information such as practices like nose picking, smoking, handwashing, handshake, and closeness to or relationship with health workers were completed by the subjects examined. The specimens were collected as follows: a sterile cotton-tipped swab was moistened in sterile distilled water and swirled inside the anterior nares and rotated clockwise and anticlockwise four times and plated by streaking onto mannitol salt agar (MSA) (Oxoid), a selective medium for the isolation of S. aureus. The plates were incubated at 37°C for 24 hours and observed for growth. After growth, staphylococci were identified on the basis of colonial characteristics, Gram stain and biochemical reactions, namely, catalase test, coagulase test (both slide and tube methods), urease test, DNAase test with DNAse agar plate (Oxoid Ltd., Basingstoke, U.K.), alkaline phosphatase test, β-galactosidase test and by fermentation of carbohydrates such as glucose, sucrose, lactose, mannitol, mannose and trehalos. Pure cultures of isolates were preserved on tryptone soy agar (Oxoid) slants for further analysis. Bacillus species were characterized by conventional microbiological methods using morphology of vegetative cells, shape and position of spores, nitrate reduction, degradation of starch, urea, casein, gelatin, acid production from glucose, mannitol, xylose, citrate utilization, lecithinase test, growth at 4°, 10°, 25°, 30°, 37°, 40°, 50°, 55°C, growth in nutrient broth with 6% NaCl [15].

#### Antimicrobial susceptibility testing

The agar diffusion method recommended by the CLSI [16] was employed to determine the susceptibility to antimicrobial agents. The following antibiotic discs were tested: ciprofloxacin,  $5\mu$ g; penicilin G, 10g; gentamicin,  $10\mu$ g; amoxycilin,  $25\mu$ g; rifampicin,  $5\mu$ g; tetracycline,  $10\mu$ g; vancomycin,  $30\mu$ g; erythromycin,  $5\mu$ g; chloramphenicol,  $30\mu$ g, augmentin,  $30\mu$ g. Zone diameters were measured after incubation for 24 hours at  $37^{\circ}$ C and results were classified following CLSI [16] interpretive standards. For the determination of methicillin resistance, the cefoxitin disk test [16] was employed. The medium used was Mueller – Hinton agar (Oxoid) and the inoculum of isolates was standardized to 0.5 MacFarland turbidity.

#### **Enterotoxin production test**

The SET-RPLA kit made by Oxoid, UK was employed to determine the enterotoxin production of the isolates of *Staphylococcus aureus* and coagulase-negative staphylococci, according to manufacturer's instructions.

#### Statistical analysis

The data obtained by questionnaires and experiment were analysed using SPSS software, version 15.0 [17]

## RESULTS

A total of 103 strains of bacteria were isolated from the 100 nasal samples collected. A total of 95 out of 100 students were

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found to be carriers of *Staphylococcus* species. Out of those positive for staphylococci 51 (53.7%) were carriers of *S. aureus*. Four of those that carried *S. aureus* also carried coagulase-negative staphylococci (3 with mixed culture of *S. epidermidis* and one with mixed culture of *S. saprophyticus*). The CONS isolated included *S. epidermidis* (11 strains), *S saprophyticus* (30 strains), *S. haemolyticus* (5 strains) and *S. saccharolyticus* (2 strains). The carriage rate of CONS among the subjects was 48%. The prevalence of MRSA among the *S. aureus* isolates was 25.5% (Table No.1). Five samples yielded *Bacillus* spp. as pure cultures, which included *Bacillus subtilis* (4 isolates) and *Bacillus cereus* (1 isolate).

The antimicrobial susceptibility of the MRSA and MRCONS is shown in Table No.2. The susceptibly of the S. aureus isolates showed that the methicillin-resistant coagulase-negative strains were more susceptible to the antibiotics tested when compared with the MRSA isolates. Only for chloramphenicol and vancomycin were higher resistance recorded for MRCONS than for MRSA. Majority of the MRSA isolates were resistant to penicillin (96.9%), amoxycillin (84.6%) and augmentin (76.9%).

Result of the enterotoxin production showed that 8 of the 13 MRSA produced staphylococcal enterotoxin A (SEA) while some of the other *S. aureus* strains produced SEA (16 strains) and SEC (5 strains). The CONS, particularly *S. saprophyticus* produced SEA (11 strains), SEB (2 strains) SEC (2 strains).

 Table No.1: Number of isolates resistant and sensitive to methicillin

Organism	Methicillin resistant	Methicillin sensitive	Total
Staph. aure	<i>us</i> 13	38	51
CONS	11	36	47
Bacillus spr	<b>)</b> . 1	4	5
Total	25	78	103

 Table No. 2: Antibiotic resistance pattern of methicillin

 resistant S. aureus and coagulase- negative staphylococci.

	% resistant	
Antibiotic	Staphylococcus aureus	Coagulase-nagative staphylococci
Penicillin	92.3	81.8
Vancomycin	0	9.1
Ciprofloxacin	46.2	18.2
Rifampicin	23.1	18.2
Augmentin	76.9	63.6
Amoxycillin	84.6	81.8
Erythromycin	46.6	45.5
Gentamicin	53.8	27.3
Tetracycline	76.9	36.4
Chloramphenico	1 15.4	36.4

#### DISCUSSION

Among the 100 students investigated, 51 (51%) were carriers of *S. aureus*. The carriage rate is higher compared with previous data in the United States [18] and among New York State prisoners [19].

It was found in this study that the ages of those who had nasal carriage of S. aureus fell mostly between ages 19 and 21 (58%). Twenty- five S. aureus were obtained from this group and of these 8 strains (32%) were MRSA strains. Prakash et al. [20] also reported 38% of S. aureus within the age group of 21-30 years with 21 isolates being methicillin resistant. However in investigations involving lower age groups in Taiwan and Turkey, a much lower percentages of MRSA were reported [6, 5]. Ogunzkaya-Artan et al. [21] reported 18% nasal carriage of S. aureus and 5.6% of MRSA among healthy preschool children. In this study 23.4% of the coagulase-negative staphylococci were resistant to methicillin, a result similar to that of Kalsoom et al.[7] and Guchi et al. [22] where 22% and 23.5% of CONS were resistant to methicillin respectively. The MRSA carriage rate 13/51 S. aureus isolates (25.5%) and among the students investigated (13%) were higher than those reported in previous data from studies from Italy [23], Trinidad and Tobago [24] and Canada [25].

The antibiotic susceptibility profiles observed in this study indicated that the MRSA were not only resistant to methicillin but to a range of other antibiotics tested.

For example resistance to penicillin was highest, 92.3% followed by amoxycillin, 84.6%, followed by augmentin and tetracycline, 76.9% each, ciprofloxacin and erythromycin (46.3% each). The least resistance was observed for chloramphenicol, 15.4%. The percentage resistance of MRSA to ciprofloxacin and erythromycin were higher than those reported by Prakash et al. [20] but lower for vancomycin. None of the MRSA in this study was resistance to vancomycin. In this study, it was observed that 16.7% of the health care workers children were carriers of MRSA. One practice of the student examined, i.e., cleaning of nostrils showed a close relationship to nasal carriage of MRSA with a P value of 0.069 ( $\chi^2$ = 3.302, df= 1). Since there were no identified risk factors in this study it was difficult to link our community isolates of MRSA to any risk factor. Some studies [26, 27] have linked recent use of antibiotics to colonisation with communityacquired MRSA but our study could not validate this by reason of the subjects examined.

There had been reports by some authors of the production of enterotoxins by coagulase- negative *Staphylococcus* isolated from humans and animals [28, 29]. In this study, majority of the enterotoxin-producing *S. aureus* produced staphylococcal enterotoxin A. The production of enterotoxins by strains of *S. aureus* is of public health significance because of their role in diarrhoea, sinusitis, sepsis and osteomyelitis [30, 31]. SEA has been reported as the most common staphylococcal enterotoxin involved in food intoxication outbreak all over the world. Enterotoxin production by coagulase–negative staphylococci should also not be ignored as CONS are becoming increasingly important as pathogens.

The results of this investigation suggests that nasal carriage of *Staphylococcus aureus* continue to play a significant role in the pathogenesis of community-acquired infections and underscores the need for continued surveillance to control emerging infections associated with MRSA. Nasal carriage of MRSA and CONS

producing enterotoxin could also add to the burden of staphylococcal infections like food poisoning, sinusitis, septic arthritis [29, 30, 32, 33].

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#### REFERENCES

1. Naimi TS, LeDell KH, Como-Sabetti K, Borchardt SM, Boxrud DJ, Etienne J, Johnson SK, Vandenesch F, Fridkin S, O'Boyle C, Danita RN, Lynfield R. Comparison of communityand health care-associated methicillin resistant *Staphylococcus aureus* infection *J. Amer.Med. Assoc.* 2003: 290: 2976-2984.

2. Saxena S, Singh K, Taiwar V. Methicillin – resistant *Staphylococcus aureus* prevalence in the east Delhi area. *Jpn. J. infect Dis* 2003: 56: 54–56.

3. Salgado CD, Farr BM, Calfee DP. Community-acquired methicillin resistant *Staphylococcus aureus*: a meta-analysis of prevalence and risk factors. *Clin. Infect. Dis.* 2003:36:131-139.

4. Ochoa TJ, Mohr J, Wanger A, Murphy JR, Heresi GP. Community-associated methicillin resistant *Staphylococcus aureus* in paediatric patients. *Emerg. Infect. Dis* 2005:11: 966-968

5. Weng-Tsung L, Wei- jen I, Min-Hua T, Jang – Jih I, Shih –Yi L, Mong- Ling C, Chih–Chien W. Nasal carriage of a single clone of community–acquired methicillin-resistant *Staphylococcus aureus* among kindergarten attendees in northern Taiwan. *BMC Infect. Dis.* 2007:7:51

6. Ciftci IH, KoKen R, Bukulmez A, Ozdemir M, Safak B, Cetinkaya Z. Nasal carriage of *Staphylococcus aureus* in 4-6 age group in healthy children in Afyonkarahuar, Turkey. *Acta Paediatrica* 2007: 96: 1043-1046.

7. Kalsoom FR, Zermina SF, Nareed A, Sattar A, Jamshaid AK, Bushra N. Nasal carriage of staphylococci in health care workers: antimicrobial susceptibility profile. *Pak. J. Pharm. Sci.* 2008: 21: 290-294

8. Klein E, Smith DL, Laxminarayan R. Hospitalizations and deaths caused by methicillin – resistant *Staphylococcus aureus*, United States, 1999-2005. *Emerg. Infect. Dis.* 2007:13: 1840-1846

9. Harbarth S, Francois P, Shrenzel J, Frankhauser-Rodriquez C, Hugonnet S, Koessler T, Huyghe A, Pittel D. Community–associated methicillin–resistant *Staphylococcus aureus* in Switzerland. *Emerg. Infect. Dis.* 2005: 11:962-965.

10. Al-Rawahi GN, Schreader AG, Porter SD, Roscoe DL, Gustafson R, Bryce EA. Methicilin-resistant *Staphylococcus aureus* nasal carriage among injection drug users; six year later. *J. Clin. Microbiol*. 2003: 46: 477-479

11. Lo W-J, Lin W-J, Tseng M-H, Wang S-R, Chu ML, Wang C-C. Community acquired methicillin resistant *Staphylococcus aureus* in children, Taiwan. *Emerg. Infect. Dis.* 2006:12: 1267-1270.

12. Karabay O, Otkun MT, Yavuz MT, Otkun M. Nasal carriage of methicillin–resistant and methicillin–susceptible *Staphylococcus aureus* in nursing home residents in Bolu, Turkey. *West Indian Med. J.* 2006:55: 183-187

13. Abudu L, Blair I, Fraise A, Cheng KK. Methicillin resistant

*Staphylococcus aureus* (MRSA): a community based prevalence survey. *Epidemiol. Infect.* 2001:126:351-356

14. Sa-Leao R, Sanches IS, Couto I, Alves CR, de Lencaster H. Prevalence of methicillin resistant strains among *Staphylococcus aureus* colonizing young and healthy members of the community in Portugal. *Microb. Drug Resis.* 2001:7:237-245.

15. Koneman EW, Allen SD, Janda WM, Schreckenberger PC, Winn Jr. WC. Color Atlas and Textbook of Diagnostic Microbiology. 5<sup>th</sup> ed. Lippincott Williams and Wilkins: Philadelphia, USA, 1997

16. Clinical and Laboratory Standards Institute (CLSI). Performance standards antimicrobial susceptibility testing; sixteenth informational supplement ,CLSI document 2006: M100-S16. CLSI: Wayne, PA.

17. SPSS. Statistical Package for Social Sciences, SPSS for Windows, Inc., Chicago, IL. USA. 2006.

18. Choi CS, Yin CS, Afra AA, Sakewi Z, Naing NN, Jamal F, Othman N. Nasal carriage of *Staphylococcus aureus* among healthy adults. *J. Microbiol. Immunol. Infect.* 2006: 39: 458-464.

19. Lowy FD, Aiello AE, Bhat M, Johnson-Lawrence VD, Lee MH, Burrell E, Wright LN, Vasquez G, Larson EL. *Staphylococcus aureus* colonization and infection in New York State prisons. *J. Infect. Dis* 2007:196:911-918

20. Prakash M, Rajasekar K, Karmegam N. Prevalence of methicillin resistant *Staphylococcus aureus* in clinical samples collected from Kanchipuram Town, Tamil Nadu, South India. *J. Appl. Sci. Res.* 2007: 3: 1705-1709. 2001: 126: 351-356.

21. Oguzkaya – Artan M, Baykan Z, Artan C. Nasal carriage of *Staphylococcus aureus* in health preschool children. *Jpn. J. Infect. Dis.* 2008: 61: 70-72

22. Guchu T, Yavuz T, Tokmak A, Behcet M, Karah E, Ozturk O, Egeli E. Nasal carriage of pathogenic bacteria in medical students: effects of clinical exposure on prevalence and antibiotic susceptibility. *Eur. Arch. Otorhinolaryngol.* 2007: 264: 85-88

23. Zanelli G, Sansoni A, Zanchi A, Cresti S, Pollini S, Rossolini GM, Cellesi C. *Staphylococcus aureus* nasal carriage in the community: a survey from central Italy. *Epidemiol. Infect.* 2002:129:417-420.

24. Akpaka PE, Kissoon S, Swanston WH, Monteil M. Prevalence and antimicrobial susceptibility pattern of methicillin

resistant *Staphylococcus aureus* isolates from Trinidad & Tobago. *Annals Clin. Microbiol. Antimicrob.* 2006: *5: 22-26* 

25. Munckhof WJ, Nimmo GR, Schooneveldt S, Schlebusch S, Stephens AJ, Williams G, Huygen F, Giffard P. Nasal carriage of *Staphylococcus aureus*, including community-associated methicillin-resistant strains, in Queensland adults. *Clin. Microbiol. Infect.* 2009:15: 149-155.

26. Bagget HC, Hennesy TW, Rudolph K, Bruden D, Reosonover A, Parkinson A, Sparks R, Donlan RM, Martinez P, Mongkolrattanothai K, Butler JC. Community onset of methicillin resistant *Staphylococcus aureus* associated with antibiotic use and cytotoxin Panton Valentine leucocidin during a furunculosis outbreak in rural Alaska. *J. Infect. Dis.* 2004: 189: 1565-1573.

27. Ellis MW, Hospenthal DR, Dooley DP, Gray PJ, Murray CK. Natural history of community-acquired methicillin resistant *Staphylococcus aureus* colonization and infection in soldiers. Clin. Infect. Dis. 2004: 39: 971-979.

28. Vernozy RC, Mazuy C, Prevost G, Lapeire C, Bes M. Enterotoxin production by coagulase-negative staphylococci isolated from goats milk and cheese. *Int. J. Food. Microbiol.* 1996: 30: 271-280

29. Udo EE. Enterotoxin production by coagulase- negative staphylococci in restaurant workers from Kuwait may be a potential cause of food poisoning. *J. Med.Microbiol.* 1990: 48: 819-823.

30. Bernstein JM, Ballow M, Schlivert PM, Rich G, Allen C, Dryja D. A superantigen hypothesis for the pathogenesis of chronic hyperplastic sinusitis with massive nasal polyposis. *Am. J. Rhinol.* 2003: 17: 321-326.

31. Mitchelin AF. Interacao das enterotoxinas estafilococicas com o sitema immune do hospedeiro. *Rev. Cienc. Farm.* 2003:24: 83-95

32. Tarkowski A, Wagner H. Arthritis and sepsis caused by Staphylococcus aureus: can the tissue injury be reduced by modulating the host's immune system? *Mol. Med. Today* 1998: 4: 15-18.

33. VonEiff C, Becker K, Machaka K, Stammer H, Peters G. Nasal carriage as a source of *Staphylococcus aureus* bacteraemia. *N. Engl. J. Med.* 2001: 344: 11-16.